Hippocampal deep brain stimulation reverses physiological and behavioural deficits in a rodent model of schizophrenia

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Abstract
Subcortical dopamine system dysregulation has been suggested to underlie the positive symptoms of schizophrenia. Recent preclinical investigations and human imaging studies have proposed that the augmented dopamine system function observed in schizophrenia patients may be secondary to aberrant hippocampal activity. Thus, we posit that the hippocampus represents a novel therapeutic target for the treatment of schizophrenia. Here we provide evidence of the effectiveness of a unique approach aimed at decreasing hippocampal function in a rodent model of schizophrenia. Specifically, in a rodent model of schizophrenia, we demonstrate that ventral hippocampal (vHipp) deep brain stimulation (DBS) can normalize aberrant dopamine neuron activity and behaviours associated with positive symptoms. In addition, we provide evidence that this approach may also be effective in restoring deficits in cognitive function, often left unaltered by conventional antipsychotic medications. Therefore, we have provided initial preclinical evidence demonstrating the feasibility of hippocampal DBS as a potential novel approach for the treatment of schizophrenia.

Key words: Cognition, deep brain stimulation, dopamine, hippocampus, MAM rat, schizophrenia.

Introduction
Schizophrenia is a debilitating disease with a lifetime prevalence of about 1% (Stilo and Murray, 2010). Aberrant sub-cortical dopamine signalling has been consistently associated with positive symptoms of schizophrenia based on multiple observations, including imaging studies and the efficacy of antipsychotic medications in treating the disease (Creese et al., 1976; Laruelle and Abi-Dargham, 1999; Seeman et al., 1975; Abi-Dargham, 2004). Given that no primary pathology exists within the midbrain dopamine system of schizophrenia patients, it has been suggested that aberrant dopamine signalling may be secondary to pathology within cortical and hippocampal regions, which are known to display progressive structural and neurochemical alterations in schizophrenia. We have recently demonstrated in a rodent model of schizophrenia (for review, see Lodge and Grace, 2009) that aberrant dopamine signalling and associated behavioural hyper-responsivity to psychomotor stimulants are due to hyperactivity within the ventral hippocampus (vHipp; Lodge and Grace, 2007, 2011). This is in accord with a growing literature demonstrating that activity within distinct hippocampal subfields is increased, at rest, in schizophrenia patients (Heckers et al., 1998; Malaspina et al., 1999; Medoff et al., 2001; Molina et al., 2003; Lewandowski et al., 2005; Schoebel et al., 2009; Tamminga et al., 2010). Furthermore, these increases in hippocampal activity have been directly correlated with clinical measures of psychosis (Molina et al., 2003; Schoebel et al., 2009). Thus, we suggest that aberrant hippocampal activity is the source of the dopamine system dysregulation in schizophrenia and, furthermore, that a novel therapeutic approach for the treatment of psychosis may be achieved by directly targeting hippocampal function (Fig. 1).

Deep brain stimulation (DBS) is being increasingly utilized for the treatment of psychiatric conditions such as depression, as well as for disorders including Parkinson’s disease and epilepsy (Krack et al., 2010). The mechanism of action of DBS has yet to be conclusively demonstrated and likely differs depending on the brain region examined. Based on clinical (Boon et al., 2007) and preclinical (Wyckhuys et al., 2010) studies, DBS of the anterior hippocampus has been purported to decrease hippocampal activity. Given our hypothesis that augmented hippocampal activity underlies dopamine dependent psychosis in schizophrenia (Lodge and Grace, 2011), we posit that DBS of the vHipp will decrease hippocampal output and subsequently normalize aberrant dopamine neuron activity and behavioural deficits.

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in an established rodent model schizophrenia. Interestingly, long-term DBS of the hippocampus has been previously investigated clinically as a treatment for epilepsy and appears to be well tolerated with no observable side-effects, albeit in a pathological condition (Boon et al., 2007; Velasco et al., 2007). Here we examined the feasibility of vHipp DBS as a therapeutic approach to reverse behavioural deficits in a rodent model of schizophrenia.

Method

All experiments were performed in accordance with the guidelines outlined in the USPHS Guide for the Care and Use of Laboratory Animals and were approved by the Institutional Animal Care and Use Committee of the University of Texas Health Science Center.

Animals

Methylazoxymethanol acetate (MAM) treatments were performed as described previously (Moore et al., 2006). In brief, timed pregnant female Sprague–Dawley rats were obtained from Harlan Laboratories at gestational day (GD) 16 and housed individually in plastic housing tubs. MAM (diluted in saline, 22 mg/kg i.p.) was administered on GD 17. Control rats received injections of saline (1 ml/kg i.p.). Male pups were weaned on post-natal day (PD) 21 and housed in groups of 2–3 with litter mates until adulthood (> PD 60) at which time they were used for physiological or behavioural studies. All experiments were performed on multiple litters of MAM and saline-treated rats.

VTA dopamine neuron extracellular recordings

All rats used for electrophysiology had not undergone any behavioural testing. Male offspring (250–400 g) were anaesthetized with chloral hydrate (400 mg/kg i.p.), as this anaesthetic does not significantly depress dopamine neuron activity (Hyland et al., 2002), and placed in a stereotaxic apparatus. Anaesthesia was maintained by supplemental administration of chloral hydrate as required to maintain suppression of limb compression withdrawal reflex. A core body temperature of 37 °C was sustained by a thermostatically controlled heating pad (TCAT-2LV; Kopf Instruments, USA). Bipolar concentric stimulating electrodes (Rhodes NEX100, Kopf Instruments) were implanted into the right vHipp (A/P +5.3, M/L +5.0, D/V −7.0 mm from bregma). Extracellular glass micro-electrodes (impedance 6–14 MΩ) were lowered into the right ventral tegmental area (VTA; A/P −5.3, M/L +0.6 mm from bregma and −6.5 to −9.0 mm ventral of brain surface) using a hydraulic micro-positioner (Model 640, Kopf Instruments). The activity of the population of dopamine neurons was determined by counting the number of spontaneously active dopamine neurons encountered while making multiple vertical passes (typically six), separated by 200 μm, in a predetermined pattern to sample equivalent regions of the VTA. Spontaneously active dopamine neurons were identified with open filter settings (low pass: 50 Hz, high pass: 16 kHz) using previously established electrophysiological criteria (Grace and Bunney, 1983) and once isolated, activity was recorded for 2–3 min. High frequency DBS (isolated current pulses: 130 Hz, 0.3 mA, 0.1 ms pulse duration were provided by a Grass S88X stimulator connected to a PSIU6X photoelectric isolation unit) was applied 10 min prior to starting electrophysiological recordings and continued throughout the duration of the experiment, typically <3 h. Sham rats were treated identically with electrodes implanted and connected; however, the stimulator remained off.

High frequency stimulation results in numerous stimulus artefacts, which presents a challenge when recording spontaneously active dopamine neurons encountered while making multiple vertical passes (typically six), separated by 200 μm, in a predetermined pattern to sample equivalent regions of the VTA. Spontaneously active dopamine neurons were identified with open filter settings (low pass: 50 Hz, high pass: 16 kHz) using previously established electrophysiological criteria (Grace and Bunney, 1983) and once isolated, activity was recorded for 2–3 min. High frequency DBS (isolated current pulses: 130 Hz, 0.3 mA, 0.1 ms pulse duration were provided by a Grass S88X stimulator connected to a PSIU6X photoelectric isolation unit) was applied 10 min prior to starting electrophysiological recordings and continued throughout the duration of the experiment, typically <3 h. Sham rats were treated identically with electrodes implanted and connected; however, the stimulator remained off.

Fig. 1. Deep brain stimulation decreases aberrant ventral hippocampal (vHipp) activity, thus restoring dopamine system function. Our hypothesis of schizophrenia suggests that aberrant hippocampal activity drives the nucleus accumbens (NAc) that, in turn, inhibits the tonic activity within the ventral pallidum. A decrease in GABAergic transmission from the ventral pallidum results in an increased dopamine neuron population activity. VTA, ventral tegmental area. (Adapted from Grace et al., 2007; for review, see Lodge and Grace, 2011.)
manipulations, such as DBS (see Fig. 1). A further difficulty, associated with the high number of stimulus artefacts, occurs when analysing firing rate and burst firing of dopamine neurons. Given that we have previously demonstrated hippocampal manipulations [both activation (Floresco et al., 2003; Lodge and Grace, 2006) and inactivation (Lodge and Grace, 2007, 2008)] do not alter these parameters, we focused this study on dopamine neuron population activity which is known to be altered in MAM-treated rats and reversed by hippocampal manipulations (Lodge and Grace, 2007; Gill et al., 2011). Electrophysiology data were analysed by a two-way analysis of variance (ANOVA; MAM and DBS as factors), followed by a Holm–Sidak post hoc test. The effects of VP manipulation were only performed in saline-treated rats and analysed by a one-way ANOVA.

**Survival surgeries**

All survival surgical procedures were performed under general anaesthesia in a semi-sterile environment. Briefly, male rats were anaesthetized with pentobarbital (60 mg/kg i.p.) and placed in a stereotaxic apparatus using bluntatraumatic ear bars. Bipolar twisted platinum stimulating electrodes (MS303/6-B/SPC: Plastics1, USA) were implanted bilaterally in the vHipp (A/P –5.3, M/L ±5.3, D/V –7.5 mm from bregma), fixed in place with dental cement and four anchor screws. Once the cement was completely solid, the wound was sutured, the rat removed from the stereotaxic frame and monitored closely until conscious. Sham rats received identical electrode implants.

**Amphetamine-induced hyper-locomotion**

All rats were connected to an 8-channel stimulus generator (STG4008, ALA Scientific Instruments, USA) via a 4-channel commutator (SL2+2C; Plastics1). DBS (isolated current pulses: 130 Hz, 0.3 mA, 0.1 ms pulse duration) was initiated immediately prior to placing rats in an open field arena (Med Associates, USA), where spontaneous locomotor activity in the x–y plane was determined for 40 min by beam breaks and recorded with Open Field Activity software (Med Associates). Sham rats were treated identically, but the leads from the commutator to the stimulator were not connected. Following the baseline period, all rats were injected with D-amphetamine sulfate (0.5 mg/kg i.p.) and locomotor activity recorded for 40 min. Finally, rats received an additional injection of D-amphetamine sulfate (2.0 mg/kg i.p.) and locomotor activity was recorded for an additional 40 min. DBS was administered continuously for the entire duration of the behavioural experiment. Locomotor data were analysed by three separate three-way ANOVAs (MAM, DBS and time as factors), one for each of the relevant time periods (spontaneous, 0.5 mg/kg, 2.0 mg/kg), followed by a Holm–Sidak post hoc test.

**Attentional set shifting**

The attentional set shifting task (AST) was performed using a method adapted from (Lapiz and Morilak, 2006). Rats examined for cognitive flexibility had been previously examined for locomotor activity or saccharine preference (data not shown). The testing apparatus was a rectangular arena divided into three quadrants. One was the start box, while the other two contained pots defined by a pair of cues along two stimulus dimensions: digging medium and odour. A Cheerio was placed at the bottom of the ‘positive’ pot and buried with the digging medium. During habituation rats were trained to reliably dig in the pots to obtain a food reward. The following day, rats were trained on a series of simple discriminations, to reach a criterion of six consecutive correct trials. Finally, on testing day MAM- and saline-treated rats were exposed to DBS (isolated current pulses: 130 Hz, 0.3 mA, 0.1 ms pulse duration) for the duration of testing (typically 3–8 h). Initially, rats were trained in a simple discrimination task of odour. Once the criterion of six consecutive trials was achieved, rats were then tested in a number of increasingly difficult tasks to include a compound discrimination, an intra-dimension shift, a second reversal, an extra-dimensional shift and a third reversal. It should be noted that during AST a subset of rats (four MAM and six saline) treated with DBS displayed seizure-like activity following long-term administration of DBS and were subsequently removed from the trial. Only data preceding the seizure were included, resulting in varying number of animals for the later stages of the task. We do not believe this to be a significant problem with the interpretation of the data since there was sufficient statistical power to identify deficits at the later stages of AST (i.e. extra-dimensional set shifting). Given that vHipp DBS is reported as an effective treatment for epilepsy, this likely reflects the need to optimize stimulus parameters. AST data were analysed by a three-way repeated measures (RM)-ANOVA (MAM, DBS and task as factors) followed by a least significant difference (LSD) post hoc test.

**Histology**

For acute/electrophysiological studies, rats were killed by an overdose of anaesthetic (chloral hydrate, additional 400 mg/kg i.p.), whereas for chronic/behavioural studies rats were killed by a lethal dose of pentobarbital (120 mg/kg i.p.). All rats were decapitated, brains removed and fixed for at least 48 h (8% w/v paraformaldehyde in phosphate buffered saline containing potassium ferrocyanide) and cryoprotected (25% w/v sucrose in PBS) until saturated. Brains were sectioned (25 μm coronal sections), mounted onto gelatin–chromium coated slides and processed with a Nissl stain for histochemical verification of electrode sites (Fig. 2). All histology was performed with reference to a stereotaxic atlas (Paxinos and Watson, 1986). It should be noted that we did not examine any histological effects of MAM.
administration as this has been performed previously with reported decreases in hippocampal area of about 15% (Moore et al., 2006).

**Analysis**

Electrophysiological analysis of single unit neuron activity was performed using commercial computer software (LabChart Pro; ADInstruments, USA), while locomotor behaviour was recorded using Open Field Activity software (Med Associates). All data are presented as the mean ± S.E.M. unless otherwise stated, with n-values representing the number of animals per experimental group. Statistics were calculated using either SigmaPlot (Systat Software Inc., USA) or Statistica (StatSoft Inc., USA).

**Materials**

MAM was purchased from Midwest Research Institute (USA); pentobarbital sodium (USP) was obtained from Lundbeck (USA). Chloral hydrate, pentobarbital sodium (non-USP), Dulbecco’s phosphate buffered saline and D-amphetamine sulfate were all purchased from Sigma (USA). All other chemicals and reagents were of either analytical or laboratory grade and purchased from various suppliers.

**Results**

**Dopamine neuron electrophysiology**

Sham rats (vHipp electrodes implanted but not stimulated) that received GD 17 saline injections (n=6 rats) exhibited an average of 1.08 ± 0.06 spontaneously active dopamine neurons per electrode track, consistent with previous findings in untreated rats (Lodge and Grace, 2007). Sham rats administered MAM prenatally (for review, see Lodge and Grace, 2009) at GD 17 (n=6) exhibited significantly greater (~2-fold) dopamine neuron population activity (1.99 ± 0.15 cell/track; two-way ANOVA; F_{MAM} = 21.484, F_{DBS} = 25.807, F_{MAM × DBS} = 22.533; Holm–Sidak; t = 6.514; p < 0.05; Fig. 3), again consistent with our previous findings (Lodge and Grace, 2007). DBS of the vHipp (130 Hz, 0.3 mA, 0.1 ms duration; n=6 rats) completely normalized the aberrant dopamine neuron population activity observed in MAM-treated rats to a level not significantly different from saline-treated rats (1.04 ± 0.09 cells/track; two-way ANOVA; Holm–Sidak; t = 6.823; p < 0.05), while vHipp DBS did not significantly alter dopamine neuron population activity in saline-treated rats (n=7 rats: 1.05 ± 0.07 cells/track; two-way ANOVA; Holm–Sidak; t = 0.240; p > 0.05; Fig. 3).

To confirm that increases in population activity could be observed during DBS, we chemically inactivated the VP in saline-treated rats. Consistent with previous observations (Floresco et al., 2003), VP inhibition significantly increased dopamine neuron activity (n=4 rats: 1.72 ± 0.10 cells/track; one-way ANOVA; F = 22.495; Holm–Sidak; p < 0.05; Fig. 3 insert) in DBS-treated rats when compared to both sham (t = 5.777, p < 0.05) and DBS only, saline-treated rats (t = 6.250, p < 0.05).

**Amphetamine-induced hyper-locomotion**

Sham, MAM-treated rats displayed a significantly enhanced locomotor response to low dose (0.5 mg/kg i.p.)

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**Fig. 2.** Histological localization of electrode placements. A representative photomicrograph depicting bilateral electrode locations within the ventral hippocampus is depicted in (a). The group data for bilateral (b) and unilateral (c) implantations are included schematically. A representative photomicrograph depicting cannula location within the ventral pallidum (VP) is shown in (d) while the group data are shown in (e). DBS, Deep brain stimulation; MAM, methylazoxymethanol acetate.
amphetamine administration when compared to sham, saline-treated controls (three-way ANOVA of 0.5 mg/kg dose; $F_{\text{MAM}}=23.462$, $F_{\text{DBS}}=6.136$, $F_{\text{MAM}} \times \text{DBS}=19.233$; Holm–Sidak $t=6.450$, $p<0.05$; Fig. 4), without changes in baseline activity (three-way ANOVA of baseline; $F_{\text{MAM}} \times \text{DBS}=0.372$; $p>0.05$) or in response to the higher dose (three-way ANOVA of 2.0 mg/kg dose; $F_{\text{MAM}} \times \text{DBS}=2.551$; $p>0.05$) consistent with previous observations (Lodge and Grace, 2007). This augmented locomotor activity was only observed with low dose amphetamine administration, as this is likely more sensitive to changes in impulse-dependent dopamine neuron activity. Consistent with the effects on dopamine neuron activity (Fig. 3), MAM-treated rats administered vHipp DBS no longer demonstrated an enhanced locomotor response to amphetamine when compared to saline-treated rats administered DBS (three-way ANOVA of 0.5 mg/kg dose; Holm–Sidak $t=0.328$; $p>0.05$; Fig. 4). These data confirm that the beneficial effect of DBS on dopamine neuron activity translates into a normalization of aberrant dopamine-mediated behaviours in a rodent model of schizophrenia. It should be noted that DBS augmented amphetamine-induced locomotor activity in saline-treated rats following the 0.5 mg/kg dose only (three-way ANOVA; Holm–Sidak $t=4.629$; $p<0.05$; Fig. 4).

**Attentional set shifting**

MAM-treated rats displayed deficits in reversal learning and extra-dimensional set shifting when compared to saline-treated rats, consistent with previous observations (Featherstone et al., 2007; Gastambide et al., 2012; three-way RM-ANOVA; $F_{\text{Task}}=12.888$, $F_{\text{Task}} \times \text{DBS}=2.646$, $F_{\text{Task} \times \text{MAM} \times \text{DBS}}=3.432$; LSD test; $p<0.05$; Fig. 5). Saline-treated rats administered DBS displayed aberrant reversal learning as demonstrated by an increase in the number of trials to meet criteria during the first reversal task (three-way RM-ANOVA; LSD test; $p<0.05$; Fig. 5). Interestingly, the same treatment that disturbed function in saline-treated animals actually normalized reversal learning in MAM-treated rats. Thus, MAM-treated rats with vHipp DBS displayed similar cognitive flexibility as saline-treated rats, with a complete reversal of the
extra-dimensional set shifting deficits and a similar trend to normalize deficits in reversal learning (three-way RM-ANOVA; LSD test; \( p < 0.05 \); Fig. 5). Taken together these data suggest that the benefits of vHipp DBS appear to be not simply limited to attenuating positive symptoms, but may also be effective at treating cognitive dysfunction.

Discussion

Increasing evidence from clinical and preclinical studies suggests that the augmented dopamine system function in schizophrenia may be secondary to aberrant hippocampal activity (Medoff et al., 2001; Lodge and Grace, 2007, 2011; Schobel et al., 2009; Heckers and Konradi, 2010). Thus, we suggest that the hippocampus represents a potential novel therapeutic target for the treatment of psychosis in schizophrenia patients (Lodge and Grace, 2011). To examine this hypothesis, we employed a rodent model of schizophrenia, namely MAM GD17, which displays deficits consistent with those observed in schizophrenia patients, including anatomical changes (deficits in parvalbumin expression and subtle reductions in the volume of medial prefrontal cortex and hippocampus), behavioural deficits (decreased prepulse inhibition of startle, stimulant induced hyper-locomotion and deficits in cognitive flexibility) and altered neuronal information processing (hippocampal and dopamine neuron hyperactivity; for review, see Lodge and Grace, 2009). Here we utilized this model to demonstrate that DBS of the vHipp can normalize aberrant dopamine neuron activity and associated behaviours. Furthermore, the beneficial effects of DBS are not limited to behaviours associated with positive symptoms, but also appear to normalize aberrant cognitive flexibility. To the best of our knowledge, this is the first experimental evidence examining the feasibility of DBS as a potential novel therapeutic approach for the treatment of schizophrenia.

As detailed earlier, augmented dopamine system function in schizophrenia is thought to be mediated by aberrant hippocampal activity (Lodge and Grace, 2007, 2011). We have previously demonstrated that the vHipp can selectively regulate the number of spontaneously active dopamine neurons in the VTA, thought to regulate the gain of the dopamine system (Floresco et al., 2001, 2003; Lodge and Grace, 2006). Specifically, activation of the vHipp leads to an increase in the number of spontaneously active dopamine neurons without altering their firing rate or burst firing pattern (Floresco et al., 2001, 2003; Lodge and Grace, 2006). This effect is likely mediated by a pathway including the nucleus accumbens (NAc) and VP (Fig. 1), as it is blocked by accumbal glutamatergic antagonist administration and mimicked by VP inactivation (Floresco et al., 2001, 2003). Similarly, MAM-treated rats display an increase in the number of spontaneously active dopamine neurons without altering their firing rate or burst firing pattern (Floresco et al., 2001, 2003; Lodge and Grace, 2006). This effect is likely mediated by a pathway including the nucleus accumbens (NAc) and VP (Fig. 1), as it is blocked by accumbal glutamatergic antagonist administration and mimicked by VP inactivation (Floresco et al., 2001, 2003). Similarly, MAM-treated rats display an increase in the number of spontaneously active dopamine neurons without altering their firing rate or burst firing pattern (Floresco et al., 2001, 2003; Lodge and Grace, 2006). This effect is likely mediated by a pathway including the nucleus accumbens (NAc) and VP (Fig. 1), as it is blocked by accumbal glutamatergic antagonist administration and mimicked by VP inactivation (Floresco et al., 2001, 2003).
provide a novel therapeutic approach for the treatment of schizophrenia. Indeed, here we provide initial evidence that vHipp DBS can normalize aberrant dopamine neuron population activity in MAM-treated rats without observable effects on dopamine system function in saline-treated animals. The finding that vHipp DBS did not alter dopamine neuron activity in control animals is consistent with our previous observations that tetrodotoxin-inactivation of the vHipp does not alter dopamine system function (Lodge and Grace, 2007, 2008) and likely reflects the low-spontaneous activity of vHipp pyramidal neurons under ‘normal’ conditions (Jung et al., 1994; Lodge and Grace, 2007).

Given the technical challenges associated with recording spontaneously active neurons during high frequency electrical stimulation (i.e. 130 stimulus artefacts per second) it is possible that the differences seen with DBS may be due to technical limitations rather than to a therapeutic effect. We do not believe this to be the case as dopamine neuron population activity was unchanged by DBS in saline-treated animals (Fig. 3) and dopamine neurons can be clearly isolated during DBS (Supplementary movie). Nonetheless, to confirm that increases in population activity could be observed during DBS, we chemically inactivated the VP, a manipulation that increases dopamine neuron activity downstream of the hippocampus (Floresco et al., 2003) and should therefore not be affected by DBS (Fig. 1). VP inhibition was able to significantly increase dopamine neuron activity in saline-treated rats receiving vHipp DBS. These data were included, not to inform on the mechanisms underlying the effects of DBS, but rather to demonstrate that the effects of DBS on dopamine neuron population activity are not simply associated with potential technical limitations of the study.

As a correlate for the beneficial effects of vHipp DBS on dopamine neuron activity we examined whether DBS also reversed behavioural deficits analogous to those observed in schizophrenia patients. To assess the effectiveness of DBS against positive symptoms, we examined the hyper-responsivity to psychomotor stimulants, which is consistently observed in both animal models and schizophrenia patients (Laruelle et al., 1996; Moore et al., 2006; Lodge and Grace, 2007). It has been suggested that an enhanced sensitivity to psychomotor stimulants is associated with an enhanced baseline dopamine neuron population activity secondary to vHipp hyperactivity (Lodge and Grace, 2007, 2011). Thus, an increase in dopamine neuron activity would be expected to augment impulse-dependent dopamine release induced by amphetamine-mediated inhibition of the dopamine transporter. Consistent with this hypothesis, MAM-treated rats display an enhanced response to low-dose amphetamine (Moore et al., 2006; Lodge and Grace, 2007). This augmented response was abolished by DBS such that there were no significant differences in the response to amphetamine in saline- or MAM-treated rats. It should be noted that DBS augmented amphetamine-induced locomotor activity in saline-treated rats, an effect that was not observed in MAM-treated rats. DBS alone did not alter dopamine neuron activity and therefore the effects of DBS in saline-treated rats may be mediated by inputs to the NAc arising from regions such as the prefrontal cortex; however, this requires further investigation. Taken together, these results demonstrate that vHipp DBS effectively normalizes aberrant dopamine neuron activity and the hyper-responsivity to psychomotor stimulants in a rodent model of schizophrenia.

Given that long-term hippocampal DBS is well tolerated in human studies (Boon et al., 2007; Velasco et al., 2007), our preliminary evidence suggests that DBS may provide a novel therapeutic approach to treat the positive symptoms of schizophrenia. It is important to note, however, that schizophrenia patients not only display positive symptoms, but also cognitive symptoms that are arguably as debilitating and not as effectively treated with conventional antipsychotics (Meltzer and McGurk, 1999). The hippocampus also regulates prefrontal cortical function (Thierry et al., 2000); therefore, it is possible that hippocampal pathology may also contribute to cognitive deficits observed in patients (Meltzer and McGurk, 1999). Thus, we examined the effects of DBS on cognitive symptoms using an AST paradigm (Lapiz and Morilak, 2006).

Consistent with previous observations, MAM-treated rats display deficits in both reversal learning and extradimensional set shifting. Hippocampal DBS was able to completely reverse these deficits in MAM-treated rats, suggesting that DBS may also be effective at treating cognitive deficits in schizophrenia patients. Extra-dimensional set shifting is known to be dependent on medial prefrontal cortical (mPFC) function in rodents (Birrell and Brown, 2000). Thus, the deficits in extradimensional set shifting observed in the MAM model likely reflect aberrant mPFC function. This could be attributable to either a direct pathology within the mPFC (for example, a loss of parvalbumin; Lodge et al., 2009), or aberrant activity of afferent inputs (i.e. from the vHipp; Lodge and Grace, 2007). Moreover, extensive literature demonstrating a relationship between dopamine and PFC performance (Floresco and Magyar, 2006; Vijayraghavan et al., 2007), suggests that the aberrant dopamine transmission observed in MAM-treated rats may also contribute to deficits in extra-dimensional set shifting. Thus, the ability of vHipp DBS to reverse deficits in extra-dimensional set shifting are likely due to either attenuation of aberrant vHipp-mPFC transmission or restoration of dopamine system function; however, the exact mechanisms remain to be elucidated.

While extra-dimensional set shifting likely involves mPFC regions, reversal learning is more often associated with orbitofrontal cortex (OFC) function (McAlonan and Brown, 2003). Interestingly, the OFC does not receive a strong hippocampal projection, suggesting that the
deficits in reversal learning observed in the MAM-treated rats are likely associated with either a direct pathology within the OFC or upstream alterations in OFC transmission (i.e. aberrant dopamine system function). It should be noted that the OFC receives reciprocal projections from the entorhinal cortex (Ongur and Price, 2000) and given that DBS can activate axonal fibres, combined with the considerable input to the hippocampus from the entorhinal cortex, it is possible that DBS may indirectly alter OFC activity. Indeed, vHipp DBS produced a deleterious effect in saline-treated rats with a significant impairment of reversal learning observed. This deficit was only observed during the first reversal task, as this task appears to be more sensitive to manipulation (Lapiz-Bluhm et al., 2009). In contrast, the opposite response was observed in MAM-treated rats, (i.e. a beneficial effect of DBS on reversal learning) possibly attributable to a normalization of aberrant dopamine neuron activity.

Taken as a whole, here we provide the first experimental evidence demonstrating the feasibility of vHipp DBS to reverse aberrant dopamine neuron activity and behaviours associated with positive symptoms in a rodent model of schizophrenia. In addition, we provide evidence that this approach may also be effective in restoring deficits in cognitive function, often left unaltered by conventional antipsychotic medications. Given that long-term DBS of the anterior hippocampus has been previously investigated in human patients and is tolerated with minimal side-effects (Boon et al., 2007; Velasco et al., 2007), we provide initial evidence that DBS may present as a novel therapeutic approach for the treatment of schizophrenia. Moreover, we provide rationale for the specific targeting of the anterior hippocampus in this disease.

Supplementary material

For supplementary material accompanying this paper, visit http://dx.doi.org/10.1017/S1461145712001344

Acknowledgements

The authors thank Dr David Morilak and Julianne Jett for their help and expertise with the attentional set shifting task. This work was supported by the NIH (R01: MH090067) and a NARSAD award from the Maltz Family Foundation.

Statement of Interest

D. J. L. received consulting fees from Dey Pharmaceuticals.

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